

Induction and Local Rules for Physics of Medicine Building

Welcome to the Physics of Medicine building (PoM)

- The Physics of Medicine Building is the focus of biological work in the Department of Physics.
- This document lays down the Local Rules for PoM. The Guidelines within are the minimum standard of work within PoM and will neither override any Risk Assessment requiring higher standards of work, nor be a substitute for the proper training required to carry out any experimental work safely.
- Failure to follow the Local Rules or specific rules for a piece of work is potentially grounds for removal of access to PoM.
- The building contains a range of laboratory spaces including CL3, CL2, CL1 and biologically undesignated laboratory areas. You must work in an area suitable for your experimental work.
- All POM lab announcements should be made to pom-daytoday@lists.cam.ac.uk. You MUST subscribe to this email list at <https://lists.cam.ac.uk/mailman/listinfo/pom-daytoday>.
- Any user removing themselves from the mailing list will be assumed to be requesting removal of access to POM.
- The latest version of this document will be made available online on the BBS website at <https://www.bss.phy.cam.ac.uk/internal/forms/>. Check regularly for updates to this document and the changes to the Local Rules that may have occurred.
- Access to PoM also requires attending and passing the Departmental H&S general health and safety induction course.
- The following courses are commonly needed for members of this department. Where relevant to your work, you should sign up for and attend the following health and safety courses found at <https://www.phy.cam.ac.uk/intranet/hands/training2> :

-  Pressurised Gases and Cryogenics
-  Chemical Safety
-  Nanoparticle Safety
-  Laser Safety for Class 3B and 4 Laser Users
-  Using Containment Facilities and Microbiological Safety Cabinets

- All work must be risk assessed BEFORE starting.
- Working out of hours requires adherence to departmental policies. There is an electronic sign-in facility, located on the ground floor near the lift.
- All users will operate to the University's Safe Biological Practise Policy as a minimum standard: [University Safe Biological Practice HSD028B Policy](#)

General Laboratory Features and Rules

- The Fire Assembly point is in front of the building on JJ Thomson Avenue and marked with the number 1. This is shared with the Maxwell building.
- A number of doors in POM have had Dorgards fitted. These red units on the base of a door allow a door to be held open if needed, but will automatically release the door if the fire alarm sounds, slowing the spread of any potential fire. If there is no Dorgard or it is not working, contact Tim

Fitzmaurice. **Do not leave any POM doors propped open with anything other than a Dorgard.**

- Toilets are located in the middle of the ground and first floor corridors, outside the laboratories.
- There is a 'No laboratory gloves on doors' policy in PoM. Gloves are disposable items, please treat them as such and remove them when moving between areas.
- Do not wear PPE you have just used in labs in non-working spaces (corridors, toilets etc). If you need to move samples etc between labs in these spaces, carry them in suitable racks or boxes.
- Eye wash facilities (the yellow plastic capped hose) are located next to every hand wash basin.
- First aid kits are located in Room 0.10 (Mechanics Lab), Room 1.08 (Chemistry Lab) and on the ground floor opposite the front entrance. If you use any items, you must notify the Laboratory Manager (Tim Fitzmaurice) or the central First Aid staff.
- All users must comply with all signs and written instructions! This is essential to keep you, your colleagues, contractors and the equipment safe.
- Switch off lights and equipment when not in use, unless there are instructions stating not to.
- All overnight/weekend experiments **MUST** be clearly labelled with your name and out of hours contact details, plus possibly an experiment description and the length of time the experiment will run for. If there is a problem, security staff may need to contact you and ask for instructions.
- **All** PoM users must ensure that communal consumables are reordered when they run low in central stocks. Some areas have rotas to cover replacement of routine use of consumables. However, not all lab spaces are included in a rota and any rotas in place are to cover routine usage, not one person's large experiment.
- Requests for replacement items should go to the Lab Manager by email or through the PoM option in this order form on the BSS website; <https://www.bss.phy.cam.ac.uk/internal/neworder>
- If you are working in any biological laboratory in PoM then by default you **MUST** wear a lab coat as a minimum of PPE due to the range of hazardous chemicals, GM agents, infectious agents or other biological agents permitted in assorted areas of PoM. Further or altered PPE requirements will be in risk assessments that have been approved by the Departmental Committees.
- **DO NOT** eat, drink, chew gum or apply cosmetics. **DO NOT** wear gloves or lab coats from other buildings. **DO NOT** bring personal/outside items into lab spaces, use hooks where provided. **DO NOT** remove lab coats from the building for use elsewhere. **DO NOT** use mobile phones with gloves in laboratories. **DO NOT** wear open toed footwear while working in laboratories.
- You **MUST** not bring any CL3 materials to non-CL3 areas nor any CL2 Biolab/Bacteria lab materials into lower category areas.
- You should avoid bringing any CL2 room PPE into CL1 areas to maintain separation of work.
- Do not remove pipettes or reagents from lab benches without checking that they are communal! Please return communal items to their location afterwards.
- Do not use any piece of equipment without training nor without permission if required.
- If you are not affiliated with BSS but require access any computers in POM, you will need to contact Will Alcock (wja28@cam.ac.uk) for access.
- The list of contacts below are the first people outside of your lab group to ask for help in the first instance. If in doubt of anything in PoM – please ask for help.

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Important Contact Details

Tim Fitzmaurice (Laboratory Manager & Biological Safety Officer): 748914 | 07722 163970 (mobile) | tjf11@cam.ac.uk | Office: Room 1.04, POM building

Saba Alai (Departmental Safety Officer): 37397 | 07817602858 (mobile) | sa792@cam | Office: Room 250, Bragg building

Anya Howe (Chemical Safety Officer & Research Support): 37397 | 07834319547 (mobile) | ajh246@cam | Office: Room 251, Bragg building

Administrators: 37478 | Room 988 & 989, Rutherford Building | rutherford.hub@phy.cam

Will Alcock (Computing Officer): 37012 | Room 906, Rutherford Building | wja28@cam

Facilities (e.g. maintenance, heating, lighting issues) | facilities.helpdesk@phy.cam

Card access: access@phy.cam

Graham Jack (BOC cylinder operative) | 65309 | Mobile: 07768558722 | jack.graham@boc.com

Out of Hours Work

Normal working hours are from 8am to 7pm on week days. Any time out of these hours, at weekends or bank holidays is officially 'out of hours'.

1. Users must sign in and out of the Out of Hours system if working between the hours of 7pm-8am or at weekends. The electronic sign-in facility is located on the ground floor near the lift. The record is automatically wiped each morning at 9am.
2. You can instead sign in and register online at: <http://lw1.cta.phy.private.cam.ac.uk/ooih/> but you must LOG OUT when you leave.
3. High risk lab work (such as work with piranha solution) is only permitted between 8:30 and 16:45, with the out of hours "buddy" system in place (CORE HOURS).
4. If carrying out Medium Risk lab work, you must work with a buddy close enough in the lab to hear you and help you if there is a problem.
5. When working in pairs using the "buddy" system, each worker must be aware of what the other person is working with or on and, importantly what to do if there is an accident. If the "buddy" is working out of sight then arrange to check on each other e.g. every 15 minutes. Arrangements for the "buddy" system must be made **before** you start relevant work. If out of hours, on arrival if you notice others are already signed IN as out of hours working, then you should inform them of your presence. If finishing medium or high risk experiments and/or leaving the laboratory, inform the other person. If there are only two persons working then inform the "buddy" of your intention to leave well before you go. This will enable the "buddy" to complete or leave his/her work in a safe state. Both of you should leave together.
6. Lone working in the PoM laboratory is acceptable only if the activity is quantified to be 'Low Risk', by formal risk assessment. You must make a copy of the risk assessment available for inspection by the Lab Manager. It is strongly recommended that you inform a colleague or the Lab Manager when you plan to work alone out of hours and when you leave the lab.

Hazardous Chemical Waste Disposal

Chemical waste is considered to be hazardous, if it contains chemicals classified as:

-  Toxic
-  Corrosive
-  Irritants
-  Harmful
-  Carcinogenic
-  Prescription only medicine
-  Flammable liquids with a flash point $\leq 60^{\circ}\text{C}$ (i.e. organic solvents)
-  Oils

You MUST carry out a (or read the relevant) hazardous substance risk assessment BEFORE you use any hazardous chemicals in PoM.

Hazardous chemical waste of different types must be handled and then stored separately. This is to prevent adverse or unexpected chemical reactions.

Disposal via the Drains

The drains are not meant for disposal of waste chemicals. Most waste chemicals will damage the waterways and the Department may be prosecuted by the Environment Agency.

However, the following can be disposed of to the drains with large volumes of water:

- Aqueous solutions containing less than 0.01% organic solvents (excluding chlorinated solvents)
- Dilute acid, alkali or ammonia solutions
- Harmless soluble inorganic salts (including CaCl_2 , MgSO_4 , NaSO_4 , P_2O_5)
- Hypochlorite solutions
- Bleach and other household cleaning agents in small amounts

DNA binding dyes are specifically banned from any drain or sink waste by Anglian Water at any level. This classification includes not just the older, highly toxic ethidium bromide compounds, but all more modern and safer alternatives as well.

There is a specific route for disposal of these dyes and any waste contaminated with them in the Biochemistry Lab (1.10) that must be listed in any Risk Assessment using them. You MUST use this route for these compounds.

For all other hazardous chemical waste, including glass or consumables that are significantly contaminated with hazardous chemicals, refer to the reference tables below.

For any queries about Chemical Waste, please see the Chemical Safety Officer (Anya Howe) or Laboratory Manager (Tim Fitzmaurice).

Biohazardous Waste Disposal

- **Solid biohazardous waste** - double bag and autoclave before disposing into the blue bins in room 1.22. Before autoclaving, store biohazardous waste in the grey bins labelled 'Un-Autoclaved Waste Only' in room 1.22.
- **Liquid biohazardous waste** - inactivate by adding Virkon to a 2% concentration and leave for a minimum of 10 minutes, before disposing down the sink with large volumes of water.

- **Biohazard contaminated gloves** - put in the grey bins labelled 'Contaminated Gloves Only' in the Cell Culture Facility.
- **Biohazard contaminated serological pipettes and pipette tips** - put in the white Bio-bins. When full, close the bin, add autoclave tape and place in the grey bins labelled 'Un-Autoclaved Waste Only' in room 1.22.
- **Biohazard contaminated needles, sharps and broken glass** – Can go straight into the sharps bins. Large quantities of biohazardous broken glass must be autoclaved before disposal in the glass bin. Ask Tim Fitzmaurice (tjf11@cam.ac.uk) for advice if unsure.
- **Used STARLAB pipette tip boxes and wafers** - put in the grey bin labelled 'Contaminated STARLAB pipette tip boxes and wafers' in room 1.22. They will then be autoclaved before recycling.

Non-hazardous Laboratory Waste

You can put the following in the regular blue waste bins, unless significantly contaminated with hazardous chemicals:

- Gloves, rinsed plasticware, paper towels/tissues etc.

Cardboard and polystyrene boxes

Ground floor labs: Leave in Mechanics lab, in the plastic tray under the bench nearest the entrance to the corridor.

First floor labs: Leave in the Biochemistry Lab, in the plastic tray under the middle bench holding the drying oven. If large amounts of cardboard need disposing of, please take to the recycling bins outside.

Collection of empty glass Winchester bottles

Ground floor labs: Leave in the wire bottle carrier racks in the Goods In room (room 0.02)

First floor labs: Leave in the wire bottle carrier racks in the Chemical Store (room 1.30)

These will be collected every month and taken to the Chemical Waste Store.

Useful Links:

BSS Equipment list: <https://www.bss.phy.cam.ac.uk/internal/bssequipment>

POM Equipment Booking: <http://people.bss.phy.cam.ac.uk/interactive/equipment>

Physics Department Health, Safety and Environment website:

<https://www.phy.cam.ac.uk/intranet/hands>

Physics Department Health, Biological work webpages:

http://www.phy.cam.ac.uk/internal_resources/hands/hazards/biology

Physics Stores online catalogue:

https://www.phy.cam.ac.uk/intranet/services/stores/Stores_CATALOGUE_Feb16.pdf

Biological Safety Induction and Human Tissue Act Training

If you are planning to work with more controlled material including malaria, blood, bacteria, algae or mammalian cells or plan to introduce any other novel biological material, please email Biological Safety Officer Tim Fitzmaurice (tjf11@cam.ac.uk) to arrange a relevant induction.

Additionally, if you are planning to carry out any work with material containing human cells that have not divided outside of the human body (e.g. teeth, hair, saliva) , this will fall under the Human Tissue Act. You will need to undergo specialist training to work with human tissue.

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Equipment/ Laboratory	Contact	Location
Laser Writer	Eileen Nugent	Mott 419A
Zeiss upright microscope	Eileen Nugent	Mott 419A
Liquid nitrogen dispenser	Tim Fitzmaurice	Goods In, POM 0.02
Sonicator	Alessio Caciagli	Shaker Room, POM 0.04
3D printers	Ulrich Keyser	Mechanics Lab, POM 0.10
Pipette pullers (model P-2000)	Ulrich Keyser	Mechanics Lab, POM 0.10
Electrophysiology measurement systems	Ulrich Keyser	Nanopore Lab, POM 0.11
FTA 200 Dynamic Contact Angle Analyzer	Pietro Cicuta	Physics Lab, POM 0.12
0.26 Microscope	Pietro Cicuta / Erika Eiser	Laser Tweezers Room, POM 0.26
0.26 Tweezers	Pietro Cicuta / Erika Eiser	Laser Tweezers Room, POM 0.26
Hyperspectral Imaging Evaluation System	Sarah Bohndiek	Hyperspectral Lab, POM 0.28
IX73 2-Deck Inverted Microscope	Otti Croze	Microbial Swimming & Ecology Imaging Room, POM 0.33
Zeiss LM510	Samuel McDermott	Leica Confocal Room, POM 0.36
Leica TCS SP5 confocal microscope	Samuel McDermott	Leica Confocal Room, POM 0.36
Electrophysiology measurement systems	Ulrich Keyser	Nanopore Lab, POM 0.11
FTA 200 Dynamic Contact Angle Analyzer	Pietro Cicuta	Physics Lab, POM 0.12
0.26 Microscope	Pietro Cicuta / Erika Eiser	Laser Tweezers Room, POM 0.26
Multitron-Pro Package Incubators	Otti Croze	Algae Lab, POM 1.06
New Brunswick BIOFLO 110 fermentor/ bioreactor	Tim Fitzmaurice	Algae Lab, POM 1.06
Z2 Dual Threshold Cell Counter	Otti Croze	Algae Lab, POM 1.06
E.M.S. Spin coater model 4000	Viola Introini	Chemistry Lab, POM 1.08
Rotary evaporator	Tom O'Neill	Chemistry Lab, POM 1.08
Differential scanning calorimetry machine	Eugene Terentjev	Biochemistry Lab, POM 1.10
GelDoc imaging system	Lorenzo Di Michele	Biochemistry Lab, POM 1.10
Nanodrop	Lorenzo Di Michele	Biochemistry Lab, POM 1.10
Centrifuge	Tim Fitzmaurice	Biochemistry Lab, POM 1.10
pH meter	Alex Ohmann	Biochemistry Lab, POM 1.10
Thermal cycler TC512	Lorenzo Di Michele	Biochemistry Lab, POM 1.10
-80°C freezer	Tim Fitzmaurice	Culture Prep Lab, POM 1.16
Microbiological safety cabinets	Tim Fitzmaurice	Cell Culture Lab, POM 1.18
Motic inverted fluorescent microscope	Tim Fitzmaurice	Cell Culture Lab, POM 1.18
-80°C freezer	Tim Fitzmaurice	Bacteria Lab 2, POM 1.20
Static incubators	Tim Fitzmaurice	Bacteria Lab 2, POM 1.20
Waterbath	Tim Fitzmaurice	Bacteria Lab 2, POM 1.20
Centrifuge	Tim Fitzmaurice	Bacteria Lab 2, POM 1.20
Ice machine	Tim Fitzmaurice	Autoclave Room, POM 1.22

Autoclave for sterilisation	Tim Fitzmaurice	Autoclave Room, POM 1.22
Waste autoclave	Tim Fitzmaurice	Autoclave Room, POM 1.22
Microbiological safety cabinet	Tim Fitzmaurice	Bacteria Lab 1, POM 1.24
Shaking incubators	Tim Fitzmaurice	Bacteria Lab 1, POM 1.24
FTIR spectrometer	Eugene Terentjev	Instrument Lab, POM 1.28
DMA machine	John Gearing	Instrument Lab, POM 1.28
Fluorescence spectrophotometer	Tim Fitzmaurice	Instrument Lab, POM 1.28
BMG Labtech FLUOstar OMEGA microplate reader	Tim Fitzmaurice	Instrument Lab, POM 1.28
UV-Vis spectrophotometers	Tim Fitzmaurice	Instrument Lab, POM 1.28

Waste Stream	What it includes	What to do	Where does it go?
Lab Smalls	Waste commercial chemical bottles (or anything not included below)	Take a blue chemical waste box from the waste store. Complete Waste Form and email to the Chemical Safety Officer (CSO) for approval and forwarding to contractor. Place bottles into chemical waste box along with a copy of the Waste Form. Place box in chemical waste store.	Chemical Waste Store
Solvent Waste	Chlorinated and non-chlorinated Solvents	Keep these in separate plastic bottles, labelling each bottle with a supplied chemical waste sticker and place in solvent waste storage area. Do not include on Waste Form. Kept in room 1.08 Chemistry Lab.	
Acid Waste	Waste acid solutions	Small amounts: dilute to less than 10% v/v. ALWAYS ADD ACID TO WATER NOT THE OTHER WAY ROUND. WEARING FULL PPE (gloves, lab coat, face shield or goggles), carefully pour diluted acid to drain in lab, while tap is running. Run tap for a few minutes after. Large amounts (inc. commercial bottles): Treat as Lab Smalls above.	
Solid Waste	Silica, drying agents Etc.	Store in a plastic container. Seal and label with type of solid waste, Group and producer name or CRSid. Container will be disposed of by waste contractor.	
Contaminated Consumables	Gloves, tissues and plastic waste contaminated with heavy metals, nanoparticles or highly toxic material	Bag up and label with type of waste and group name. This should then be treated as Lab Smalls above.	
Sharps Waste	Metal sharps inc:- needles with syringes (glass syringe can be separated if safe to do so), scalpels and blades	Place into sharps boxes. When box is full completely seal the lid. Cover biological hazard sign with the supplied 'Chemical Sharps' sticker and label with group name. Take to the Chemical Waste Store. Container will be disposed of by waste contractor.	

Glass Waste	Any laboratory glass item (including broken glass) – does not include normal drinking glasses/bottles	Place in white waste tub for glass. Keep lid on tub when not in use.	
Empty Glass Winchesters	Clean empty chemical bottles and Winchesters	Clean all chemical bottles (if in doubt, ask the CSO), deface or rip off the label and place in mixed recycling. DO NOT PUT WASHINGS DOWN THE DRAIN. Empty Fisher Winchesters should be placed in empty Fisher cardboard box.	
Uncontaminated Consumables	Gloves, Tissues and Plastic waste Etc.	Bag up and place in domestic waste. This can be in a normal bin bag. But ensure it is sealed before cleaners arrive. Alternatively take to waste bins in the loading bay.	Cleaner pick up or take to domestic waste skip.

If in doubt: Contact the Chemical Safety Officer (CSO), Anya Howe; Email ajh246@cam.ac.uk, Tel: 37421, Mobile: 07834 319547

POM Laboratory Information

<u>Room</u>	<u>Person Responsible</u>	<u>Equipment</u>	<u>Significant Hazards</u>
0.02 Goods In	N/A	Liquid nitrogen dispenser Delivered goods are left here.	Asphyxiation, cryogenics
0.04 Shaker Room	Erika Eiser (ee247@cam.ac.uk)	Sonicator, MCR501 Modular Compact Rheometer	Noise
0.06 Hub Room	N/A		
0.08 Controlled Molecular Transport Lab	Ulrich Keyser (ufk20@cam.ac.uk)		Lasers, asphyxiation
0.10 Mechanics Lab	Ulrich Keyser (ufk20@cam.ac.uk)	3-D printers, dissection microscope, Visco Analyser V4000, plasma cleaner, micropipette puller, PDMS balances and dessicators First aid box: next to the hand basin.	Chemicals, sharps
0.11 Nanopore Lab	Ulrich Keyser (ufk20@cam.ac.uk)	Tecella Triton+ 16 channel electrophysiology measurement system	
0.12 Physics Lab	Pietro Cicuta (pc245@cam.ac.uk)	Fume hoods, dissection microscope, FTA 200 Dynamic Contact Angle Analyzer, various microscopes	Chemicals
0.13 Olympus Microscope Room	Arijit Das (akd41@cam.ac.uk)	FV1200 Olympus confocal microscope	Lasers, asphyxiation, biohazard
0.14 Malaria Lab	Pietro Cicuta (pc245@cam.ac.uk)	Biological safety cabinet, incubator, microscope, centrifuge, liquid nitrogen culture storage dewar	Asphyxiation, cryogen, biohazard
0.16 Microscopy Lab	Sarah Bohndiek (seb53@cam.ac.uk)		Lasers
0.24 Laser Interface Scattering Room	Pietro Cicuta (pc245@cam.ac.uk)		Lasers
0.26 Laser Tweezers Room	Pietro Cicuta (pc245@cam.ac.uk) / Erika Eiser (ee247@cam.ac.uk)	Two Nikon Ti-E inverted microscopes	Asphyxiation, biohazard
0.28 Hyperspectral Labs	Sarah Bohndiek (seb53@cam.ac.uk)	Hyperspectral Imaging Evaluation System	Lasers, biohazard
0.30 Cell Mechanics Lab	Ulrich Keyser (ufk20@cam.ac.uk)	FV1200 Laser scanning microscope, Eclipse TE2000U microscope	Lasers, biohazard
0.31 Cilia Lab	Pietro Cicuta (pc245@cam.ac.uk)		

<u>Room</u>	<u>Person Responsible</u>	<u>Equipment</u>	<u>Significant Hazards</u>
0.32 Laser Scattering Room	Eugene Terentjev (emt1000@cam.ac.uk)		Lasers
0.33 Microbial Swimming & Ecology Imaging Room	Otti Croze (oac24@cam.ac.uk)	IX73 inverted microscope	Lasers, biohazard
0.34 Atomic Force Microscope	Pietro Cicuta (pc245@cam.ac.uk)	Nikon Ti-E inverted microscope	Lasers, biohazard
0.35 Optics Lab	Sarah Bohndiek (seb53@cam.ac.uk)	WITEC raman and fluorescence microscope	Lasers, asphyxiation, biohazard
0.36 Leica Confocal Room	Sam McDermott (sjm263@cam.ac.uk)	Leica confocal microscope, Zeiss LM510	Lasers, asphyxiation, biohazard
1.02 Office		Lab write up area	
1.04 Office	Tim Fitzmaurice (tjf11@cam.ac.uk)		
1.06 Algae Lab	Otti Croze (oac24@cam.ac.uk)	Laminar flow cabinet, centrifuge, shaking incubator, Beckman Coulter Counter Z series	Chemicals, biohazard
1.08 Chemistry Lab	Eugene Terentjev (emt1000@cam.ac.uk)	Fumehoods, electrospinner, balances, glassware oven, spin coater, rotary evaporator This lab's glassware: in room 1.32. First aid box: on the bench next to handwash sink. Lab coats: on pegs next to handwash sink.	Chemicals; flammables, toxic, harmful, corrosive
1.10 Biochemistry Lab	Tim Fitzmaurice (tjf11@cam.ac.uk)	Differential scanning calorimetry machine, millipore filtered water, magnetic stirrers / hotplates, vortex, ultrasonic baths, GelDoc Imaging system, fume hoods, glassware oven, pH meter, centrifuges, balances, drying oven, nanodrop, microwave This lab's glassware: in room 1.28. Clean lab coats: in cupboard beneath balances.	Chemicals, biohazard

<u>Room</u>	<u>Person Responsible</u>	<u>Equipment</u>	<u>Significant Hazards</u>
		Distribution points for syringes, tissues, gloves and glass vials for whole building.	
1.12 Nucleic Acids Room	Ulrich Keyser (ufk20@cam.ac.uk)	PCR machines, laminar flow cabinet	Chemicals
1.14 Bacteriophage Lab	Diana Fusco (df390@cam.ac.uk)	Incubator	Biohazard, asphyxiation
1.16 Cell Culture Prep Room	Tim Fitzmaurice (tjf11@cam.ac.uk)	-80°C freezer, liquid nitrogen culture storage dewar	Asphyxiation, cryogen, biohazard
1.18 Cell Culture Lab	Tim Fitzmaurice (tjf11@cam.ac.uk)	Biological safety cabinets, incubators, centrifuge, inverted fluorescence microscope Lab coats: in lobby outside the room.	Asphyxiation, biohazard
1.20 Bacteria Lab 2	Tim Fitzmaurice (tjf11@cam.ac.uk)	Centrifuge, water bath, -80°C freezer, incubators Non-sterile glassware for bacteria lab is stored here.	Biohazard
1.22 Autoclave Room	Tim Fitzmaurice (tjf11@cam.ac.uk)	Ice machine, autoclaves, dishwasher	Biohazard, heat/steam from the autoclave
1.24 Bacteria Lab 1	Tim Fitzmaurice (tjf11@cam.ac.uk)	Biological safety cabinet, shaking incubators, centrifuge, microwave Sterile glassware for bacteria lab is stored here.	Biohazard
1.28 Instrument Room	Tim Fitzmaurice (tjf11@cam.ac.uk)	FTIR machine, DMA machine, FLUOstar OMEGA plate reader, Cary 300 Bio UV VIS spectrophotometers, Cary Eclipse fluorimeter	Biohazard, asphyxiation
1.30 Chemical Store	Eugene Terentjev (emt1000@cam.ac.uk)	Spare chemicals and equipment are stocked here; UV crosslinker, balances, Silverson L4R mixer, Hetofrig heater, Branson Sonifier 150, magnetic hotplate stirrer, signal generator.	Flammables, acids, toxic chemicals
1.32 Glassware Store	Eugene Terentjev (emt1000@cam.ac.uk)		Chemicals